## Total Synthesis of the 5-Methylenephosphonate Analogue of D-myo-Inositol 1,4,5-Trisphosphate

## J. R. Falck,\* Abdelkrim Abdali, and Steven J. Wittenberger

Departments of Molecular Genetics and Pharmacology, University of Texas Southwestern Medical Center, Dallas, Texas 75235, USA

The 5-methylenephosphonate analogue of p-myo-inositol 1,4,5-trisphosphate has been prepared from (-)-quinic acid and shown to be a long-lived agonist of calcium mobilization.

D-myo-Inositol 1,4,5-trisphosphate (1,4,5-IP<sub>3</sub>), the calcium mobilizing intracellular second messenger of the phosphatidylinositol (PI) cycle, is rapidly metabolized *in vivo* by either of two divergent pathways: (a) initial phosphorylation by a 3-kinase followed by hydrolysis of the C(5)-phosphate, or (b)

direct dephosphorylation by a specific 5-phosphatase.<sup>1</sup> Intervention in these pathways may provide new insights into the function and possible therapeutic manipulation of the PI cycle. In particular, it has been deemed of value to prepare inositol (poly)phosphates modified with metabolically more

 $Bn = PhCH_2$ 

OPO(OH)2

HO

(7)

Scheme 1. Reagents and conditions: i, mesyl chloride (1.5 equiv.), Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, 0°C, 20 min; ii, NaOMe (0.15 м), MeOH/THF (1:2), 0°C, 30 min; iii, PCC (1.5 equiv.), 3 Å molecular sieves, HOAc (0.2 equiv.), 4 h at 0°C, then 1.5 h at 24°C; Et<sub>3</sub>N (1.5 equiv.), 24°C, 15 min; iv, NaBH<sub>4</sub>, MeOH/THF (3:1), 0°C, 1 h, 68% overall from (1); v, SEM-Cl (2 equiv.), Pri<sub>2</sub>NEt (5 equiv.), THF, 45 °C, 12 h, 85%, vi, DIBAL-H (3.5 equiv.), PhMe, -78 °C, 3 h, 76%; vii, N-(phenylseleno)phthalimide (2 equiv.), Bu<sub>3</sub>P (2 equiv.), THF, 0°C, 45 min, 65%; viii, NaIO<sub>4</sub> (2 equiv.), 1,4-dioxane/pH 7 buffer (1:1.6), 0°C, 6 h, 85%; ix, KH (2.2 equiv.), PhCH<sub>2</sub>Br (2 equiv.), THF, 12 h, 90%; x, Bun<sub>4</sub>NF (0.25 m, 3 equiv.), HMPA, 105 °C, 65 min, 92%; xi, PhSeBr (1.1 equiv.), Na<sub>2</sub>CO<sub>3</sub> (1.4 equiv.), CH<sub>2</sub>Cl<sub>2</sub>,  $m\text{-ClC}_6\text{H}_4\text{CO}_3\text{H}$  (1.3 equiv.),  $\text{C}_5\text{H}_5\text{N}$  (2.6 equiv.),  $-78 \rightarrow 0$  °C, 0.5 h, 95% (crude); xii, NaPO(OCH<sub>2</sub>Ph)<sub>2</sub> (2.8 equiv.), 18-crown-6 (1 equiv.), PhMe, 0°C, 20 min, 65%; xiii, BH<sub>3</sub> (3 equiv.), THF,  $0 \rightarrow$ 24°C over 1.25 h, then 24°C for 1.5 h, m-ClC<sub>6</sub>H<sub>4</sub>CO<sub>3</sub>H (9 equiv.), 25 °C, 14 h, 95%; xiv, Pri<sub>2</sub>NP(OCH<sub>2</sub>Ph)<sub>2</sub> (4 equiv.), 1H-tetrazole (16 equiv.),  $CH_2Cl_2$ ,  $0 \rightarrow 24$  °C, 2 h; m-ClC<sub>6</sub>H<sub>4</sub>CO<sub>3</sub>H (6 equiv.),  $-78 \rightarrow$  $0^{\circ}$ C, 3 h, 91%; xv, 10% Pd/C, H<sub>2</sub> (3.5 atm), EtOH/H<sub>2</sub>O (4:1), 25 °C, 5 h; EtOH/H<sub>2</sub>O/HOAc (24:6:1), 24°C, 16 h; NaOH, MeOH, 67%.

stable phosphorus functionalities.<sup>2</sup> Herein, we describe the total synthesis of (7), the 5-methylenephosphonate analogue of 1,4,5-IP<sub>3</sub>, utilizing a strategy<sup>3</sup> that exploits both the chirality and complete carbon framework of commercial (–)-quinic acid.

Lactone (1), obtained<sup>4</sup> from (-)-quinic acid in 86% yield, was converted to the known<sup>3</sup> ester (2) (68% overall) by an improved procedure involving sequential mesylation of the tertiary alcohol, lactone methanolysis using NaOMe, and pyridinium chlorochromate (PCC) oxidation. Following elimination of methanesulphonate using triethylamine in methanol/tetrahydrofuran (THF) (1:2), the newly generated enone was reduced in situ by sodium borohydride exclusively from the less hindered  $\beta$ -face (Scheme 1). Protection of the C(1)-alcohol (inositol numbering) in (2) as its  $\beta$ -trimethylsilylethoxymethyl (SEM) ether (85%), low temperature di-isobutylaluminium hydride (DIBAL-H) reduction of the ester (76%), and selenylation<sup>5</sup> (65%) of the resultant primary alcohol generated selenide (3).† Stereoselective [2,3]-sigmatropic rearrangement (85%) of the allylic selenoxide derived from (3) and benzylation (90%) gave rise to (4) as the sole product.

Fluoride ion induced desilylation (92%) of (4) in hexamethylphosphoric triamide (HMPA) and kinetically controlled addition<sup>6</sup> of phenylselenenyl bromide to the exocyclic alkene provided mainly the anti-Markovnikov adduct. The latter underwent smooth oxidative elimination to the somewhat labile allylic bromide (5)‡ (95%), which was sufficiently pure to be used directly in the next step. Michaelis-Becker phosphorylation<sup>7</sup> (65%) of (5) using excess sodium dibenzyl phosphite in the presence of 18-crown-6 and hydroboration of the alkene with 3-chloroperoxybenzoic acid work-up furnished a single stereoisomer (95%) identified by <sup>1</sup>H NMR analysis (250 MHz, COSY, J-resolved) as (6)‡ with the desired all-trans configuration between the substituents on C(3)–C(6). Introduction of the C(1) and C(4) phosphates (91%) by the two-step phosphite method according to Tegge and Ballou<sup>8</sup> and removal of the protecting groups afforded (7)‡ (67%), isolated as its sodium salt.

Initial biological evaluation suggests that (7) acts as a long-lived agonist of 1,4,5-IP<sub>3</sub> activity. It elicits contraction of bovine tracheal smooth muscle permeabilized with saponin<sup>9a</sup> and stimulates a sustained release of calcium from a microsomal preparation of bovine adrenal gland.<sup>9b</sup> The details of these studies will be presented elsewhere.

This work was financially supported by a Grant-in-Aid from

‡ Selected data for (5): m.p. 143—144 °C (Et<sub>2</sub>O/hexane);  $[\alpha]_D^{23}$  $52.5^{\circ}$  (c 1.30, CHCl<sub>3</sub>);  $\delta_{\rm H}$  (250 MHz, CDCl<sub>3</sub>) 1.31—1.68 (m, 10H), 2.33 (d, 1H,  $J \sim 6.8$  Hz), 3.88 (d, 1H,  $J \sim 9.8$  Hz), 3.93 (ddd, 1H,  $J \sim 2.6, 3.0, 7.0 \text{ Hz}), 4.35 (d, 1H, J \sim 9.8 \text{ Hz}), 4.43 (d, 1H, J \sim 7.5 \text{ Hz}),$ 4.48 (ddd, 1H,  $J \sim 1.0$ , 2.6, 5.2 Hz), 4.59 (ddd, 1H,  $J \sim 1.5$ , 3.4, 5.2 Hz), 4.88 (d, 1H,  $J \sim 10.8$  Hz), 4.96 (d, 1H,  $J \sim 10.8$  Hz), 5.79 (br. s, 1H), 7.29—7.46 (m, 5H). For (6):  $[\alpha]_D^{23}$  +21° (c 1.24, CHCl<sub>3</sub>);  $\delta_H$  $(250 \text{ MHz}, \text{CDCl}_3) 1.33-1.80 \text{ (m, 1H)}, 1.85 \text{ (dddd, 1H, } J \sim 6.0, 6.2,$ 8.3, 26.9 Hz), 2.12 (dd, 1H,  $J \sim 6.0$ , 18.8 Hz), 2.30—2.55 (br. s, 2 × OH), 3.66 (dd, 1H,  $J \sim 6.2$ , 8.3 Hz), 3.92 (dd, 1H,  $J \sim 8.3$ , 11.1 Hz),  $3.95 (dd, 1H, J \sim 4.4, 6.2 Hz), 4.06 (dd, 1H, J \sim 6.3, 8.3 Hz), 4.41 (dd, 1H, 1H, 1Hz)$ 1H,  $J \sim 4.3$ , 6.3 Hz), 4.47 (d, 1H,  $J \sim 11.3$  Hz), 4.68 (d, 1H,  $J \sim 11.3$ Hz), 4.88-5.08 (m, 4H), 7.14-7.44 (m, 15H); FAB MS (m/z) 610 (M + H, 100%), 519 (M + H – PhCH<sub>2</sub>, 25%). For (7) (hexasodium salt):  $\delta_{\rm H}$  (500 MHz, D<sub>2</sub>O) 1.25 (dt, 1H, J ~10.6, 15.0 Hz), 1.57—1.66 (m, 1H), 2.05 (apparent dd, 1H,  $J \sim 15.0$ , 20.1 Hz), 3.53—3.59 (m, 2H), 3.68—3.78 (m, 2H), 4.20 (apparent t, 1H,  $J \sim 2.2$  Hz); <sup>31</sup>P NMR (81 MHz, D<sub>2</sub>O, pH 12, <sup>1</sup>H decoupled)  $\delta$  3.05 (s, 1P), 4.73 (s, 1P), 21.58 ppm (s, 1P); FAB MS (m/z) 551 (M + H, 25%), 529 (24%), 507 (12%).

<sup>†</sup> Satisfactory analytical data were obtained for all new, stable intermediates.

the American Heart Association and the Robert A. Welch Foundation.

Received, 14th February 1990; Com. 0/00685H

## References

- 1 Review: C. P. Downes, Biochem. Soc. Trans., 1989, 17, 259.
- 2 A. M. Cooke, R. Gigg, and B. V. L. Potter, J. Chem. Soc., Chem. Commun., 1987, 1525; C. E. Dreef, G. A. van der Marel, and J. H. van Boom, Recl. Trav. Chim. Pays-Bas, 1987, 106, 512; A. M. Cooke, N. J. Noble, R. Gigg, A. L. Willcocks, J. Strupish, S. R. Nahorski, and B. V. L. Potter, Biochem. Soc. Trans., 1988, 16, 992;
- C. Schultz, T. Metschies, and B. Jastorff, *Tetrahedron Lett.*, 1988, **29**, 3919; J. J. Kulagowski, *ibid.*, 1989, **30**, 3869.
- 3 Cf. J. R. Falck and P. Yadagiri, J. Org. Chem., 1989, 54, 5851.
- 4 D. Mercier, J. Leboul, J. Cleophax, and S. D. Gero, *Carbohydr. Res.*, 1971, **20**, 299.
- P. A. Grieco, J. Y. Jaw, D. A. Claremon, and K. C. Nicolaou, J. Org. Chem., 1981, 46, 1215.
- 6 P.-T. Ho and R. J. Kolt, Can. J. Chem., 1982, 60, 663.
- 7 R. Engel, 'Synthesis of Carbon-Phosphorus Bonds,' CRC Press, Boca Raton, Florida, 1988, pp. 7—21.
- 8 W. Tegge and C. E. Ballou, Proc. Natl. Acad. Sci. USA, 1989, 86,
- 9 (a) J. T. Stull (Univ. Texas Southwestern), personal communication; (b) J. Ely and K. Catt (National Institutes of Health, USA), personal communication.